$4\alpha$ ,15-Dihydroencelin (3). Colourless crystals, mp 175° (Et<sub>2</sub>O); IR  $v_{\rm max}^{\rm CCl_4}$  cm<sup>-1</sup>: 1760 (γ-lactone), 1670 (C=CC=O); MS m/z (rel. int.): 246.126 [M]<sup>+</sup> (21) (C<sub>15</sub>H<sub>18</sub>O<sub>3</sub>), 231 [M – Me]<sup>+</sup> (3), 218 [M – CO]<sup>+</sup> (3), 97 (71), 95 (79), 69 (100), 55 (98).

$$[\alpha]_{24^{\circ}}^{\lambda} = \frac{589}{+14} \quad \frac{578}{+14} \quad \frac{546}{+20} \quad \frac{436 \text{ nm}}{+53} \text{ CHCl}_3; c \ 0.1$$

Acknowledgements—We thank the Fonds der Chemie and the Deutsche Forschungsgemeinschaft for financial support.

## REFERENCES

- 1. Bohlmann, F., Zdero, C. (1977) Phytochemistry 16, 786.
- 2. Robinson, H. (1981) Smithsonian Contributions to Botany 51, 1.
- Czerson, H., Bohlmann, F., Stuessy, T. F. and Fischer, N. H. (1979) Phytochemistry 18, 257.
- Bohlmann, F., Jakupovic, J., Ates (Gören), N., Schuster, A., Pickardt, J., King, R. M. and Robinson, H. (1983) *Liebigs Ann. Chem.* 962.

- 5. Herz, W. and Kumar, N. (1979) Phytochemistry 18, 1743.
- Bohlmann, F., Dhar, A. K., Jakupovic, J., King, R. M. and Robinson, H. (1981) Phytochemistry 20, 838.
- 7. Sims, J. J. and Bergmann, K. A. (1972) Phytochemistry 11,
- Bjeldanes, L. F. and Geissman, T. A. (1971) Phytochemistry 10, 1079.
- Geissman, T. A. and Mukherjee, R. (1968) J. Org. Chem. 33, 656.
- Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1982) Phytochemistry 21, 2329.
- 11. Bohlmann, F. and Lonitz, M. (1980) Chem. Ber. 113, 2410.
- Bohlmann, F., Ziesche, J., King, R. M. and Robinson, H. (1981) Phytochemistry 20, 1623.
- Bohlmann, F. and Jakupovic, J. (1979) Phytochemistry 18, 1189
- Guerreiro, E., Kavka, J., Giodarno, O. and Gros, E. G. (1979) *Phytochemistry* 18, 1235.
- Rodriguez, E., Saunders, B., Grieco, P., Majetick, G. and Oguri, T. (1979) Phytochemistry 18, 1641.

Phytochemistry, Vol. 23, No. 5, pp. 1187-1188, 1984. Printed in Great Britain.

0031-9422/84 \$3.00 + 0.00 Pergamon Press Ltd.

## 2-ACETOXY-3α,4α-EPOXY-3,4-DIHYDROKAUNIOLIDE FROM GROSVENORIA COELOCAULIS

FERDINAND BOHLMANN, CHRISTA ZDERO, ROBERT M. KING\* and HAROLD ROBINSON\*

Institute for Organic Chemistry, Technical University of Berlin, D-1000 Berlin 12, West Germany; \*Smithsonian Institution, Department of Botany, Washington, DC 20560, U.S.A.

(Revised received 4 November 1983)

Key Word Index-Grosvenoria coelocaulis; Compositae; sesquiterpene lactones; guaianolides.

Abstract—The aerial parts of Grosvenoria coelocaulis gave the known guaianolides dehydroleucodin, desacetoxymatricarin, kauniolide and a new one,  $2\beta$ -acetoxy- $3\alpha$ ,  $4\alpha$ -epoxy-3, 4-dihydrokauniolide.

Grosvenoria (Compositae, tribe Eupatorieae) is a small genus [1] ranging from central Ecuador into northern Peru, which is placed in the subtribe Critoniinae [2]. So far nothing is known on the chemistry of this genus. We have now investigated Grosvenoria coelocaulis (B. L. Robins.) K. et R. from northern Peru. The aerial parts afforded  $\alpha$ -curcumene and  $\alpha$ -zingiberene as well as the known guaianolides kauniolide (1) [3], dehydroleucodin [4], desacetoxymatricarin [5] together with a new one, C<sub>17</sub>H<sub>20</sub>O<sub>5</sub>. The IR spectrum of the latter indicated the presence of a y-lactone and an acetate group (1770, 1738, 1245 cm<sup>-1</sup>). From the <sup>1</sup>H NMR spectrum (Table 1) the presence of 6\alpha,12-methylene lactone could be deduced. A typical fourfold doublet at  $\delta 2.80$  was due to the H-7 signal. Accordingly, its irradiation collapsed the H-13 doublets to singlets, the double doublet at  $\delta$ 3.63 to a doublet and changed the overlapped multiplet around 2.10 while a quartet at 1.33 collapsed to a triplet. Addition of deuteriobenzene allowed the assignment of all signals by spin decoupling. As H-2 showed a W-coupling with H-5 and an allylic coupling with H-14 the whole sequence leading to the structure 2 could be assigned. The chemical shifts of H-2 and H-3 in combination with the other data indicated a 2-acetoxy derivative of kauniolide where the 3,4-double bond was transformed to an epoxide. Inspection of a model showed that the small coupling  $J_{2,3}$ required a trans-orientation of H-2 and H-3, while the chemical shift of H-15 obviously was influenced by the deshielding effect of the lactone oxygen at C-6. This, however, required a  $4\beta$ -methyl group as the stereochemistry at C-5-C-7 clearly followed from the trans-diaxial couplings of H-5-H-7. Also the observed couplings of H-8 and H-9 nicely agreed with the angles which could be deduced from a model. Thus the new compound is  $2\beta$ - 1188

Table 1. <sup>1</sup>H NMR spectral data of 2 (400 MHz, TMS as int. standard)

	CDCl <sub>3</sub>	CDCl <sub>3</sub> -C <sub>6</sub> D <sub>6</sub> (2:1)
H-2	5.73 br s	5.54 ddq
H-3	3.65 br s	3.45 br d
H-5	3.33 br d	3.05 br d
H-6	3.63 dd	3.16 dd
H-7	2.80 dddd	2.34 dddd
Η-8α	2.10 m	1.68 dddd
Н-8β	1.33 dddd	0.96 dddd
Η-9α	2.29 br dd	1.94 br dd
Η-9β	2.16 m	1.82 ddd
H-13	6.15 d	5.97 d
H-13'	5.40 d	5.08 d
H-14	1.70 s	1504
H-15	1.65 s	1.50 br s
OAc	2.12 s	1.90 s

J (Hz): 2, 3 = 2; 2, 5 = 2, 14  $\sim$  1; 5, 6 = 6, 7 = 10; 5, 14  $\sim$  1; 7, 8 $\alpha$  = 3; 7, 8 $\beta$  = 11; 7, 13 = 3.5; 7, 13' = 3; 8 $\alpha$ , 8 $\beta$  = 13; 8 $\alpha$ , 9 $\alpha$   $\sim$  1; 8 $\alpha$ , 9 $\beta$  = 6; 8 $\beta$ , 9 $\alpha$  = 12; 8 $\beta$ , 9 $\beta$  = 1.5; 9 $\alpha$ , 9 $\beta$  = 14.

acetoxy- $3\alpha$ ,  $4\alpha$ -epoxy-3, 4-dihydrokauniolide (2). The chemotaxonomic relevance of such guaianolides may be shown by further investigations of *Grosvenoria* species as well as of species of other genera placed in the subtribe Critoniinae. So far sesquiterpene lactones only are reported from *Critonia* species [6, 7]. Guaianolides closely related to those isolated now were isolated from *Kaunia* species [3], a genus of the related subtribe Oxylobinae.

## **EXPERIMENTAL**

The air dried plant material (110 g, voucher RMK 9258, collected in January 1983 in northern Peru) was extracted with Et<sub>2</sub>O-petrol-MeOH (1:1:1). The extract obtained was separated first by CC (silica gel) after methanol insoluble parts were removed. The petrol fractions gave by TLC (silica gel, AgNO<sub>3</sub> coated, petrol: detection always by UV-light and spraying with KMnO<sub>4</sub>) 10 mg  $\alpha$ -curcumene ( $R_f$  0.45) and 50 mg  $\alpha$ -zingiberene ( $R_f$  0.40). The combined fractions with Et<sub>2</sub>O-petrol (1:1, Et<sub>2</sub>O and Et<sub>2</sub>O-MeOH, 10:1) were further purified by TLC (silica gel PF 254, Et<sub>2</sub>O-petrol, 3:1) leading to a crude fraction containing a methylene lactone ( $^1$ H NMR). HPLC (RP 8, MeOH-H<sub>2</sub>O, 3:2)

gave 5 mg 1 ( $R_t$  5.4 min), its <sup>1</sup>H NMR spectrum being identical with that of authentic kauniolide. A more polar fraction from the TLC contained a mixture which was not separated by HPLC (RP 8, MeOH-H<sub>2</sub>O, 3:2:  $R_t$  4.1 min). TLC (silica gel, CHCl<sub>3</sub>-C<sub>6</sub>H<sub>6</sub>-Et<sub>2</sub>O, 2:2:1) gave 15 mg dehydroleucodin ( $R_f$  0.65, identical by <sup>1</sup>H NMR and mp with authentic material), 5 mg desacetoxy matricarin ( $R_f$  0.60) (identical with authentic material by <sup>1</sup>H NMR and mp) and 5 mg 2 ( $R_f$  0.52). Quantities were determined by weight.

 $2\beta$ -Acetoxy-3α,4α-epoxy-3,4-dihydrokauniolide (2). Colourless crystals, mp 191° (Et<sub>2</sub>O). IR  $v_{max}^{CHCl_3}$  cm<sup>-1</sup>: 1770 (γ-lactone), 1738, 1245 (OAc); MS m/z (rel. int.): 304.131 [M]<sup>+</sup> (4) (calc. for C<sub>17</sub>H<sub>20</sub>O<sub>5</sub>: 304.131), 289 [M-Me]<sup>+</sup> (3), 262 [M-ketene]<sup>+</sup> (33), 261 [289 - CO]<sup>+</sup> (100), 247 [262 - Me]<sup>+</sup> (50), 244 [M-HOAc]<sup>+</sup> (56), 216 [244 - CO]<sup>+</sup> (61), 135 (95).

$$[\alpha]_{24^{\circ}}^{\lambda} = \frac{589}{+92} \frac{578}{+96} \frac{546}{+107} \frac{436 \text{ nm}}{+173} \text{ CHCl}_3; c \ 0.17$$

Acknowledgements—We thank Dr. L. E. Bishop, Seattle, Washington, for his help and the Fonds der Chemie for financial support.

## REFERENCES

- 1. King, R. M. and Robinson, H. (1975) Phytologia 30, 221.
- 2. King, R. M. and Robinson, H. (1980) Phytologia 46, 446.
- Bohlmann, F., Kramp, W., Gupta, R. K., King, R. M. and Robinson, H. (1981) Phytochemistry 20, 2375.
- 4. Bohlmann, F. and Zdero, C. (1972) Tetrahedron Letters 621.
- Holub, M. and Herout, V. (1962) Coll. Czech. Chem. Commun. 27, 2980.
- Bohlmann, F., Jakupovic, J. and Lonitz, M. (1977) Chem. Ber. 110, 301.
- Bohlmann, F., Suwita, A., Natu, A. A., Czerson, H. and Suwita, Ant. (1977) Chem. Ber. 110, 3572.